

## **Core Imaging Facility**

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## **DAPI Staining Protocol**

Prepare the following solutions:

(To determine the amount needed for your experiment, please note that one tube of **Solution 1** will yield a total of 100mL of working solution, at a final concentration of 0.5ug/mL.)

**Solution 1\*:** 5 μL DAPI (10 mg/ml solution)

995 μL PBS

\* Solution is viable for one month if stored at 4° in the dark

**Solution 2 (working solution):** 10 μL of Solution 1

990 μL PBS

**Treatment:** →Add **Solution 2** (working solution) to your tissue/cells and incubate at room temperature for 1-

5 minutes.

 $\rightarrow$ Rinse 2X with PBS.

→Mount coverslip with Shur/Mount, Prolong or comparable antifade medium.

→Seal coverslips (if necessary) with Valap or nail polish.\*\*

\*\*Try to avoid nail polish if are visualizing GFP, as the additives in nail polish tend to quench the signal.